Field Sampling Guidelines for PFAS Using EPA Method 533, 537 or 537.1

Please read entire instruction sheet prior to sampling.

Also, view the MassDEP video on how to conduct PFAS sampling at: https://youtu.be/zrwhwSI-R9M

Sampling for PFAS using EPA method 533, 537 or 537.1 can be challenging due to the prevalence of PFAS compounds in consumer products. Many materials normally used in field and laboratory operations contain PFAS such as tubing, sample containers, and sampling tools. Since these products can contain PFAS, they cannot be used in sampling for PFAS. In addition, many consumer goods, such as water-resistant jackets or fast food wrappers, brought to a sampling site may contain PFAS that can also contaminate samples.

Field Clothing and Personal Protective Equipment

- Do not wear clothing or boots containing Gore-Tex®
- Wear new nitrile gloves
- Wet weather gear should be made of polyurethane and PVC only
- Wear safety boots made from polyurethane and PVC
- Do not use materials containing Tyvek® or polytetrafluoroethylene (PTFE)
- Do not use fabric softener on clothing to be worn in field
- Do not use cosmetics, moisturizers, hand cream, or other related products the morning of sampling
- Do not use prohibited sunscreen or insect repellant. See *Do's and Don'ts* table below for more information.

Food Considerations

No food or drink allowed on-site with exception of bottled water.

Field Equipment

- Must not contain Teflon® (aka PTFE) or LDPE materials.
- All sampling materials must be made from stainless steel, HDPE, acetate, silicone, or polypropylene.
- No waterproof field books can be used.
- No plastic clipboards, binders, or spiral hard cover notebooks can be used.
- Sharpies® and permanent markers not allowed; regular ball point pens are acceptable.
- Keep PFAS samples in separate cooler, away from sampling containers that may contain PFAS.
- Coolers filled with regular ice only Do not use chemical (blue) ice packs.

Sample Containers

- All sample containers must be made of HDPE or polypropylene
- Caps must be unlined and made of HDPE or polypropylene (no Teflon®-lined caps)

Equipment Decontamination

- Have "PFAS-free" water on-site for decontamination of sample equipment. No other water sources are to be used.
- Only Alconox® and Liquinox® can be used as decontamination materials.

Sampling Protocol

If sampling for other contaminants, sample for PFAS first. Other containers for other methods may have PFAS present on their sampling containers. Use a dedicated cooler for PFAS samples.

Before sampling, in order to limit contamination, the sample handler must wash their hands and wear new nitrile gloves when filling and sealing the sample bottles.

Each sample set requires a set of three containers to comply with the method, as indicated in box. (A sample set is composed of all samples collected from the same sample site at the same time.)

For a sample set you will need:

- 3- empty 250 mL sampling bottles preserved with 1.25 g Trizma[®].
- 1-field blank (FRB) empty 250 mL bottle unpreserved.
- 1-250 mL reagent water for field blank use preserved with 1.25 g Trizma[®].

Field Blanks

Field blanks are required and allow for the identification of interferences/contamination introduced during sample collection and handling. These containers are provided by the laboratory. Field blanks must always be collected but are analyzed only if PFAS is detected in the field sample.

Field Blank Instructions

- 1. At the sampling location, locate the reagent water container from the bottle order. The reagent water container will be pre-filled with PFAS-free water that has been preserved with Trizma®.
- 2. Locate the empty bottle labeled "field blank."
- 3. Handle the bottles as detailed in **Sampling Instructions** below. Open the bottles and transfer contents of the preserved reagent water container into the field blank bottle.
- 4. The Field blank needs to be noted on the Chain of Custody (COC).
- 5. Both the <u>empty</u> reagent water container and the <u>filled</u> field blank bottle must be returned to the lab in the cooler along with the samples taken.

Sampling Instructions

Samples collected from active drinking water sources should be collected during normal operating conditions.

- 1. Each sampling event requires three containers (one sample and two field duplicates).
- 2. Before sampling, remove faucet aerator.
- 3. Avoid contact with any Teflon® tape or pipe thread paste on pipe fittings or sampling tap threads on the water supply discharge pipe.
- 4. Run water for five minutes to flush, slow water to flow of pencil thickness to avoid splashing when filling.
- 5. Open sample bottle; do not place the cap on any surface and avoid all contact with the inside of the sample bottle and its cap.
- 6. Fill sample bottle to neck of bottle and do not overfill (or allow preservative to escape.)
- 7. After collecting the sample, cap the bottle securely and agitate by hand (approx. 5 times) until preservative is dissolved. Do not re-open bottle from this point forward.
- 8. Place each bottle in a sealed ZipLoc® bag, and place in a cooler that only contains PFAS samples in it. (No other sample types allowed.)
- 9. Ensure COC and all labels on bottles contain the required information. The samples, field blank and empty reagent blank containers are to be in the ice-filled cooler (do not use blue ice packs) and returned to the laboratory. Samples should be kept at 4°C ±2. Samples must not exceed 10°C during first 48 hours after collection. Hold time is 14 days.

Do's and Don'ts

Detection of PFAS at very low levels can be influenced by materials that are present at the sampling site, materials used by the sample collector, or sample container handling practices. The following table provides a summary of items that are likely to contain PFAS and therefore should not be used by the sample collector at the sampling site.

| Category | Prohibited Items | Allowable Items |
|--|---|---|
| Pumps and Tubing | Teflon®, polytetrafluoroethylene (PTFE) and other fluoropolymer containing materials | High-density polyethylene (HDPE), low density polyethylene (LDPE) bladders , silicone tubing, or peristaltic pump or stainless steel submersible pump |
| Decontamination | Decon 90 | Alconox ${\mathbb R}$ or Liquinox ${\mathbb R}$, potable water followed by deionized rinse |
| Sample Storage and Preservation | LDPE or glass bottles, PTFE-or Teflon®-lined caps, chemical ice packs, and waterproof labels | Laboratory-provided sample container -preferred; or, HDPE or polypropylene bottles, regular ice, bubble wrap, passive diffusion bags, and hydrasleeves |
| Field Documentation | Waterproof/treated paper or field books, plastic clipboards, spiral bound notebook, Sharpie® and permanent markers | Plain Paper, metal clipboard, pens, ZipLoc® resealable plastic storage bags, aluminum foil, Post-It® and other adhesive paper products, water-level tape |
| Clothing | Clothing or boots made of or with Gore-Tex™ or other synthetic water resistant and/or stain resistant materials including polyethylene fiber suits, Tyvek® material, waterproof boots | Synthetic or cotton material, previously laundered clothing (preferably previously washed greater than six times) without the use of fabric softeners |
| Personal Care Products (on day of sample collection) | Cosmetics, moisturizers, hand cream, sunscreen, insect repellants and other related products | Sunscreens: Alba Organics Natural, Yes to Cucumbers, Aubrey Organics, Jason Natural Sun Block, Kiss My Face, Baby-safe sunscreens ('free' or 'natural') Insect Repellents: Jason Natural Quit Bugging Me, Repel Lemon Eucalyptus, Herbal Armor, California Baby Natural Bug Spray, BabyGanics Sunscreen and Insect Repellents: Avon Skin So Soft Bug Guard-SPF 30 |
| Food and Beverage | Pre-packaged food, chemical ice packs, fast food wrappers or containers | Resealable plastic storage bags, aluminum foil, bottled water or hydration drinks |

 ${\it Please contact MassDEP Drinking Water Program with additional questions or concerns.} \\ {\it Program.Director-DWP@mass.gov}$